

# Respiratory Specimens: From collection to clinical decision-making

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**MBChB (UCT), DTM&H (Wits), FC Path (SA) (Microbiology), MMed (Microbiology) (Wits)**



## OVERVIEW

1. Specimen selection & collection
2. Basic concepts (MC&S)
  - Microscopy/Culture/Sensitivity
3. Lower respiratory tract cultures
4. Molecular Diagnostic Tests
5. Mycobacterial Diagnostics
6. Blood cultures
7. CRP
8. PCT





# 1. Specimen selection & collection

At a basic level, the doctor needs answers to 3 very basic questions from the laboratory:

1. Is my patient's illness caused by an infection?
2. If so, what microbe (bacteria, virus, parasite, fungus)?
3. What is the susceptibility profile of the organism so therapy can be targeted?



**A properly selected & collected specimen is the single most important step in the diagnosis of any infectious disease**

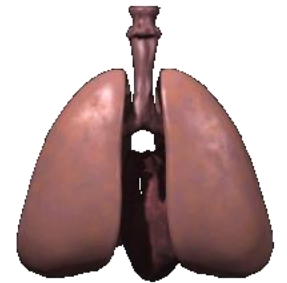
General rules for all specimens:

1. The **quantity** of the specimen must be adequate
2. It should be **representative** of the infectious process
3. **Avoid** specimen **contamination** by using sterile containers & observing aseptic technique
4. Take samples **before** administering **antibiotics**
5. **Prompt transportation** to the laboratory



# Lower Respiratory Tract Infections

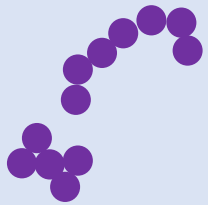
- Respiratory tract infections are among the most common infectious diseases
- The list of causative agents continues to expand as new pathogens & syndromes are recognised
- **Clinical microbiology is a science of interpretive judgement that is becoming more complex, not less**
- Key points for the laboratory diagnosis of lower respiratory tract infections:
  - PCRs have largely replaced rapid antigen tests & culture for respiratory virus detection
  - First morning expectorated sputum is always best for bacterial culture
  - Blood cultures may occasionally be helpful, particularly in hospitalised patients with pneumonia
  - In the immunocompromised host, a broad diagnostic approach based upon invasively obtained specimens is suggested
  - Bronchoscopy with washings is the optimal diagnostic specimen in paediatrics





## 2. MC&S Basic Concepts – Microscopy (Gram Stain)

Gram +



e.g. *Staphylococcus aureus*



Gram -



e.g. *Klebsiella pneumoniae*



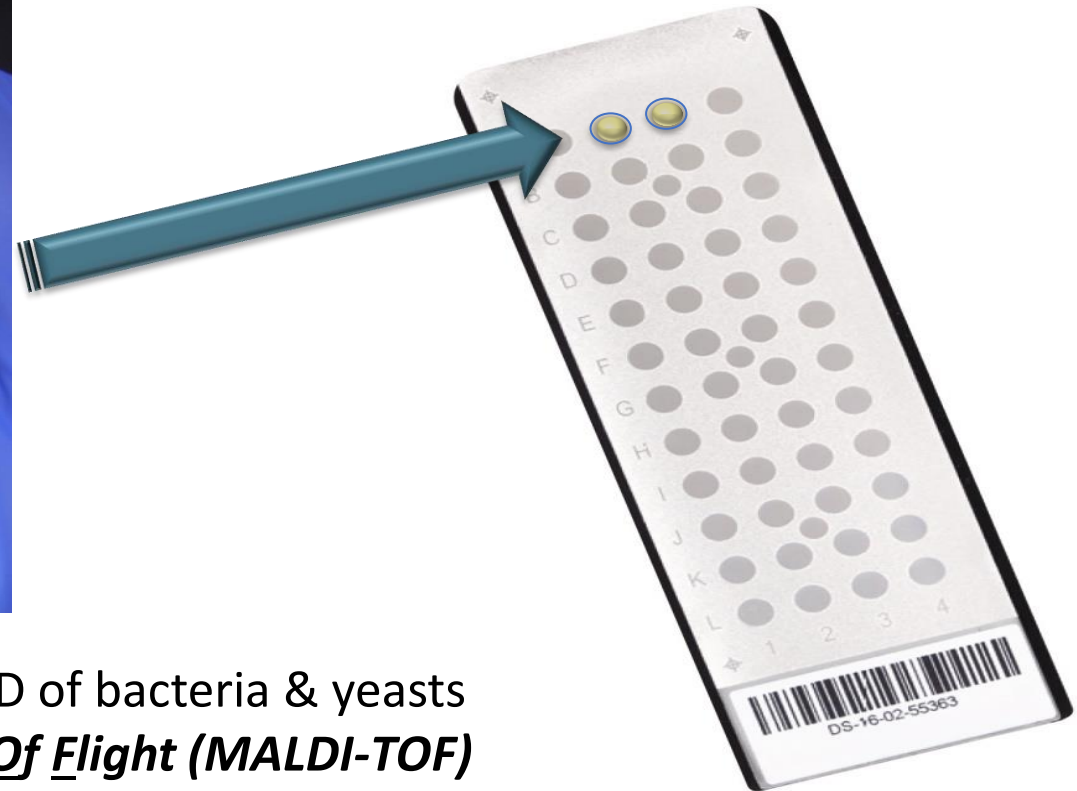
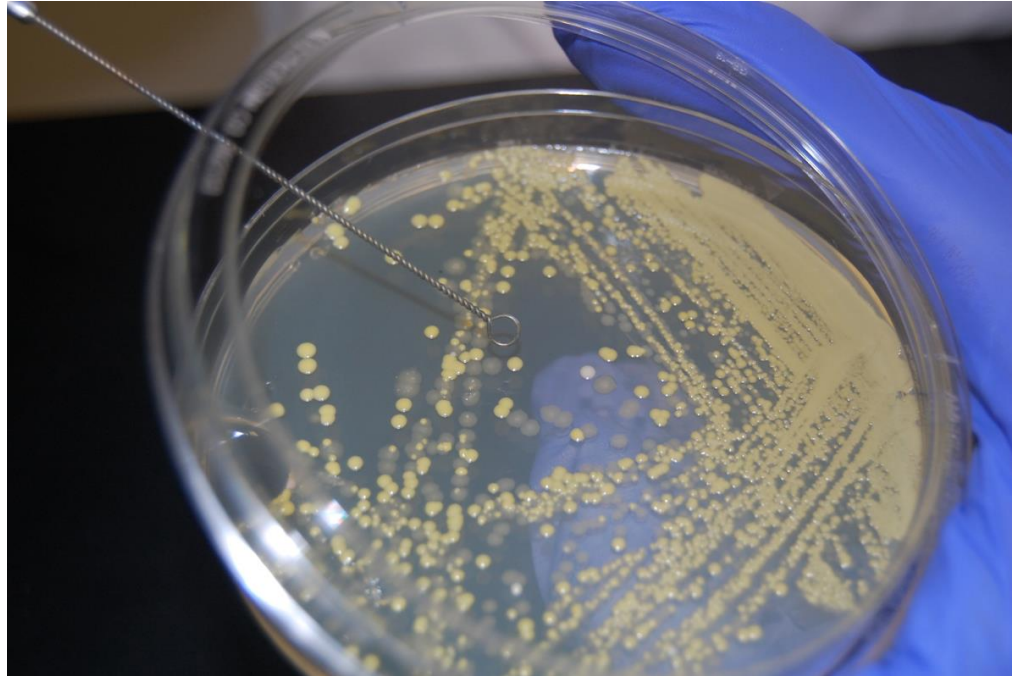


# Basic Concepts - Culture Media





# Basic Concepts – Organism Identification



One of the most important advances in post-culture ID of bacteria & yeasts is ***Matrix-Assisted Laser Desorption Ionization-Time Of Flight Mass Spectrometry (MS)***

Based on ID of characteristic protein patterns



# MALDI-TOF MS



Biomérieux Vitek<sup>®</sup> MS



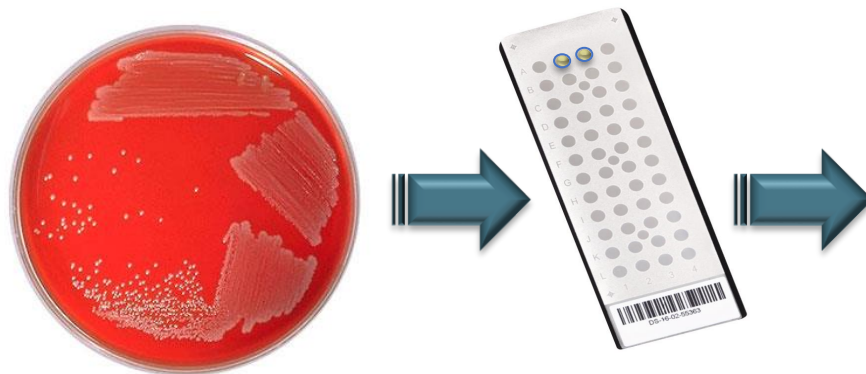
Bruker MALDI Biotyper<sup>®</sup>



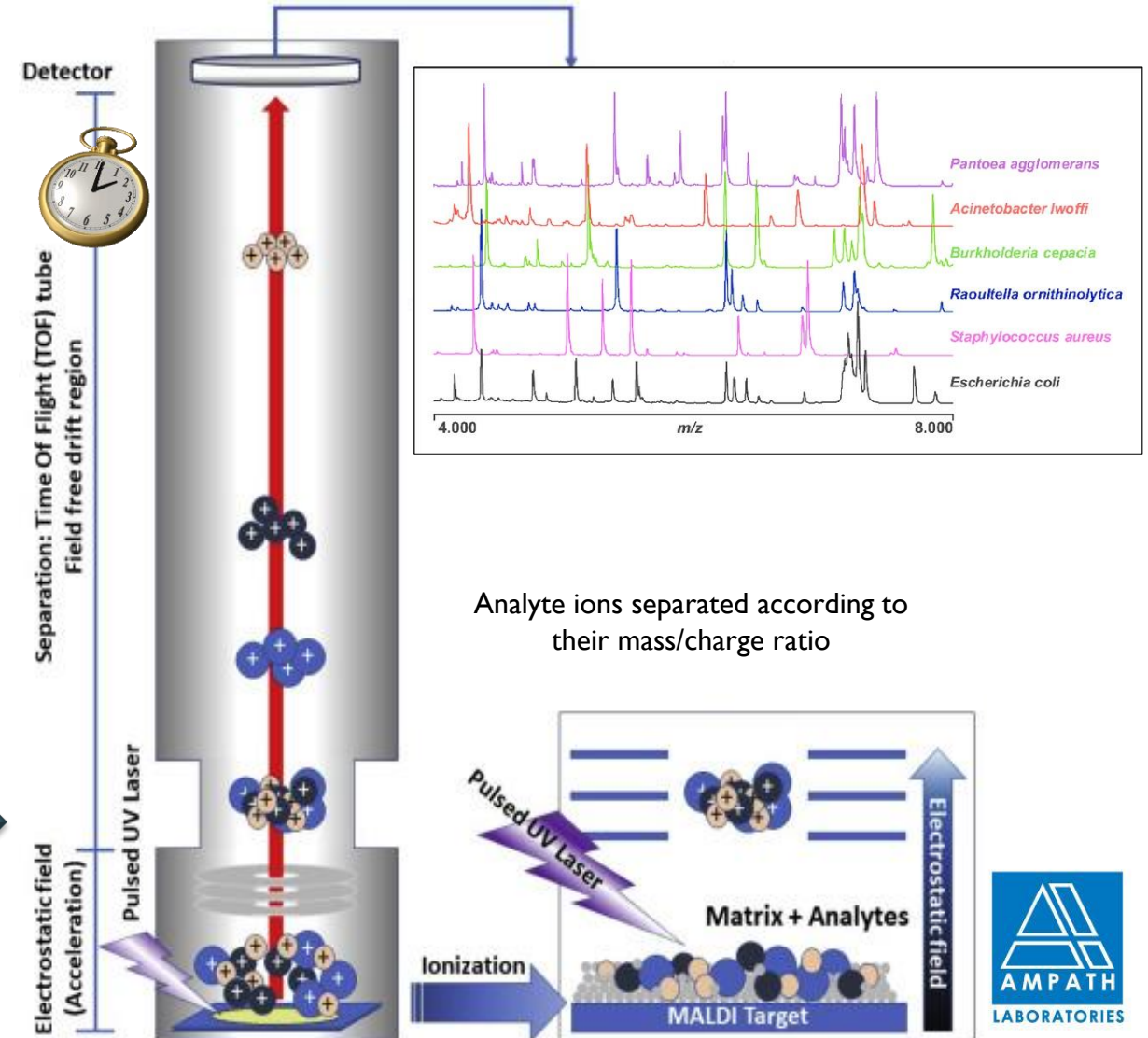


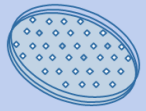
# Principle of MALDI-TOF MS

- A pulsed UV-laser is used to evaporate the matrix & the analyte into the gas phase
- In addition to desorption support, the matrix ionizes the analyte molecules by proton transfer
- The ions are then accelerated by an electrical field & enter the field-free flight tube of the mass spectrometer
- By simple measurement of the TOF of the analyte ions, their mass to charge ratio can be determined



Singhal N, et al. Front Microbiol 2015;6:791.



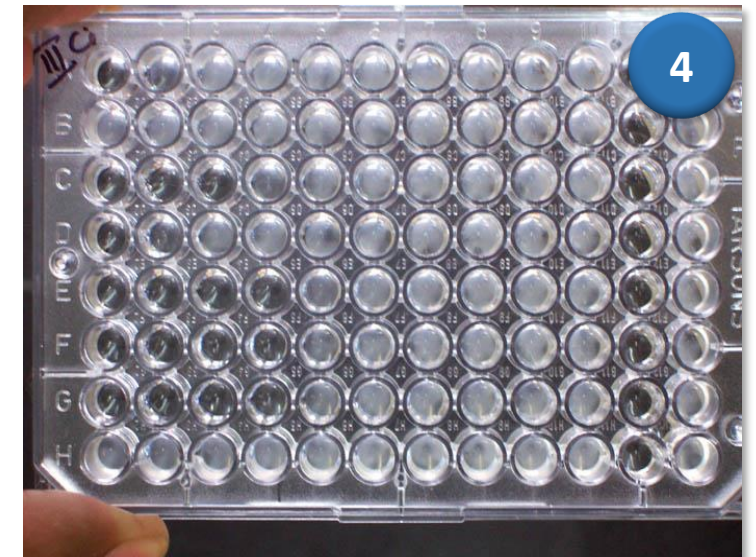
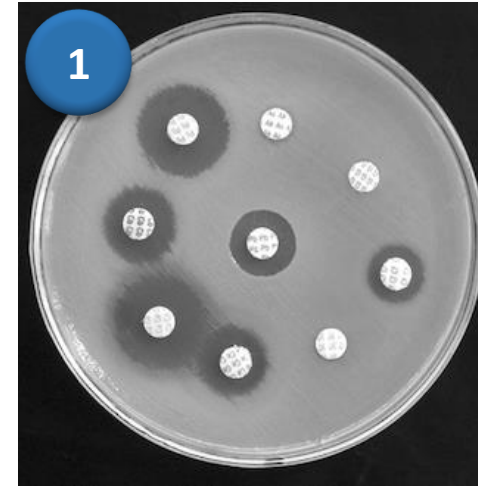


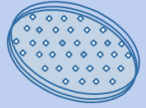
# Basic Concepts - Antimicrobial Susceptibility Testing (AST)

 AST can be carried out using a variety of methods


 The methods most commonly used are:

1. Disk diffusion (Kirby-Bauer)
2. Antibiotic gradient strips (Etest<sup>®</sup>)
3. Automated systems (Vitek<sup>®</sup>)
4. Broth microdilution






# Basic Concepts - Minimum Inhibitory Concentration (MIC)

 **MIC is the lowest concentration of the antibiotic that results in inhibition of visible growth** (*i.e.* colonies on a plate or turbidity in broth culture)

- under standard conditions
- with a standard inoculum

 The MIC is the main pharmacodynamic parameter that quantifies susceptibility against an antibiotic

 Categorise bacteria as:

- S = Susceptible
- I = Susceptible, increased exposure (*i.e.* higher dose)
- R = Resistant





### 3. Lower Respiratory Tract Specimens

■ Interpretation of culture results must take the following into consideration:

1. **QUALITY** of the specimen on microscopy
2. Quantitation of growth (heavy, moderate, or light)
3. **ALWAYS** correlate clinically and radiological for significance

■ Rates of true pathogen detection vary among studies:

- Good quality specimens: 73% (95% CI 26-96%)
- Lower quality specimens: 36% (95% CI 22-53%)

■ ***Candida* spp. does NOT cause pneumonia**

■ *Candida* spp. is a respiratory coloniser



**Important!**



Good Quality

vs.

Bad Quality



# Interpretation of Lower Respiratory Tract Cultures

## 1. QUALITY of the specimen on microscopy

- Few/no polymorphic white cells & many epithelial cells (from mucous membrane of the mouth) indicate specimen consists of or contains saliva  
➔ **culture results will therefore not be reliable** ➔ **send repeat specimen**
- Numerous polymorphs with only scanty epithelial cells & culture results that correlate with Gram stain ➔ **indicate a 'good' specimen**
- The presence of >25 epithelial cells indicates a sputum with oropharyngeal contamination and therefore grounds for rejection for bacterial culture



## 1. Quantitation of growth (heavy, moderate, or light)

- True pathogens are generally present in moderate or heavy amounts

## 3. ALWAYS correlate clinically and radiological for significance

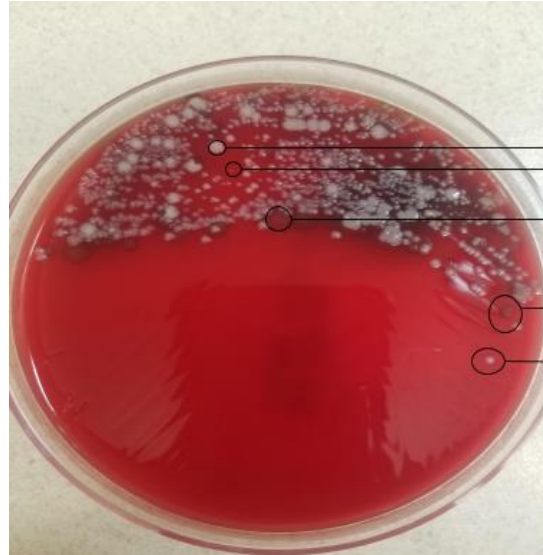


Bartlett's Grading System for Assessing Quality of Sputum Specimens	
No. of neutrophils per 10X low power field	Grade
< 10	0
10 – 25	+ 1
> 25	+ 2
Presence of mucus	+ 3
No. of epithelial cells per 10X low power field	Grade
< 10	0
10 – 25	- 1
> 25	- 2





# Example: Sputum culture



Few growth of at least 5 different colony types:

- (1) Large white
- (2) Small clear
- (3) Large spreading alpha
- (4) Smaller coining alpha
- (5) smaller white

**Blood agar**

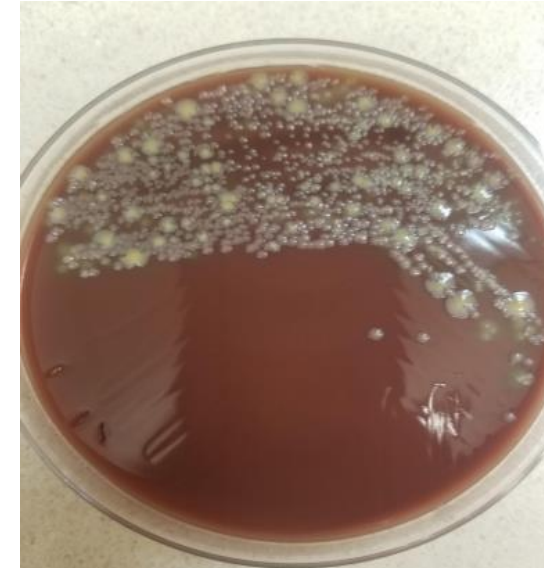


Rare lactose-fermenting Gram negative rod

Rare pinpoint non lactose-fermenting Gram negative rod

Box added to cover identifying information

**MacConkey agar**



Same as blood agar: few growth of 5 or more colony types

**Chocolate agar**



# Sputum Cultures

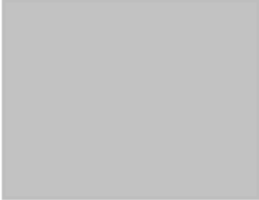


Drs Du Buisson, Kramer, Swart, Bower Inc.  
Registration number: 2007/018337/21  
PR0520005200431

## Pathology Report

24hr Contact No. 011 709 1000

Patient:



Doctor:

**Dr T Law**  
8 Stuzdee Avenue  
Ground Floor  
Rosebank, JHB 2021



**FINAL REPORT**

Req: [Redacted]  
Specimen: 20:RE00378175

Collected: 2020-10-10 01:07  
Received: 2020-10-11 01:07  
Printed: 2021-11-17 10:26  
Batch: Email 436671 I-STD

Copy Doctor: Ward Solomon Stix Morewa Rehab Ward, Solomon Stix Infection Control  
Ordered: Sputum MCS, Sputum microscopy comment  
Comments: No collection date/time on the request form  
Sputum MCS

### PROCEDURE

Specimen: Sputum

### >MACROSCOPY

Appearance : Mucoid

### >MICROSCOPY

Pus cells : < 10 cells / LPF  
Squamous epithelial cells : > 25 cells / LPF  
Gram pos cocci : Numerous  
Gram neg bacilli : Numerous  
Gram neg cocci : Numerous  
Gram neg coccobacilli : Moderate  
Yeast cells : Absent

### >MYCOBACTERIUM MICROSCOPY

Acid fast bacilli Absent

**2. Quantitation of growth**

### >AEROBIC CULTURE

Acinetobacter nosocomialis : Scanty  
Growth : Final  
Id sens : Final

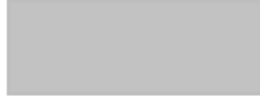
**1. Quality of specimen**



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PR0520005200431

## Pathology Report

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Doctor:

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Batch: Email 436671 I-STD

### >AEROBIC CULTURE

	A.noso
<b>BETA-LACTAMS</b>	
Ceftazidime	R
Cefepime	R
Imipenem	R
Meropenem	R
Doripenem	R
<b>AMINOGLYCOSIDES</b>	
Amikacin	R
Gentamicin	R
<b>QUINOLONES</b>	
Ciprofloxacin	R
<b>OTHER</b>	
Colist/Polymyx	S
CoL/Poly MIC 2	S
Cotrimoxazole	R
Tigecycline	S

### >COMMENT

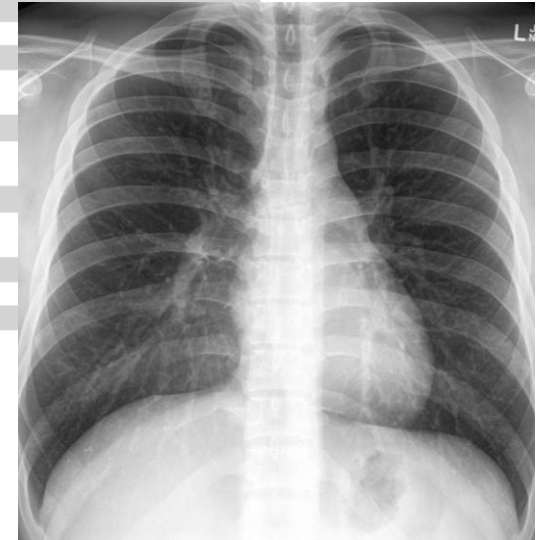
Acinetobacter nosocomialis

Acinetobacter nosocomialis  
This is an extensively drug-resistant isolate. Strict isolation, contact precautions and dedicated nursing of colonised and/or infected patients are advised. Please contact your local microbiologist regarding possible treatment options or if further advice is needed.

### >COMMENT

The presence of >25 epithelial cells per LPF indicates a sputum with probable oropharyngeal contamination.

**3. Correlate with CXR**



**No new infiltrates on CXR  
Coloniser!**





# Tracheal Aspirate Culture

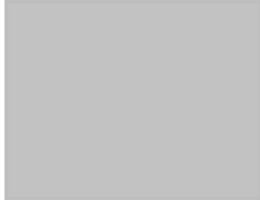


## Pathology Report

Drs Du Buisson, Kramer, Swart, Bower Inc.  
Registration number: 2007/018337/21  
PR0520005200431

24hr Contact No. 011 709 1000

Patient:



Doctor:

Dr T Law  
8 Sturdee Avenue  
Ground Floor  
Rosebank, JHB 2021

FINAL REPORT

Req: [Redacted]

Specimen: 21:RE00289985

Collected: 2021-07-23 14:00  
Received: 2021-07-23 15:55  
Printed: 2021-11-17 10:39  
Batch: Email 436692 I-STD

Copy Doctor: Dr T Law, Rosebank Clinic Infection Cont, Rosebank Clinic ICU

Ordered: Tracheal MCS, Fungal culture, Carbapenemase Cult Screening, Pneumocystis jirovecii PCR, Mycobacterium PCR

### PROCEDURE

Specimen: Tracheal aspirate

### RESULT

Description:

#### >MACROSCOPY

Appearance : Mucoïd

1. Quality of specimen

#### >MICROSCOPY

Pus cells : > 25 cells / LPF  
Squamous epithelial cells : < 10 cells / LPF  
Gram neg bacilli : Numerous  
Yeast cells : Absent

2. Quantitation of growth

#### >MYCOBACTERIUM MICROSCOPY

Acid fast bacilli : Absent

#### >AEROBIC CULTURE

Klebsiella pneumoniae : Profuse

#### >COMMENT

Klebsiella pneumoniae

This isolate tested positive for carbapenemase production with PCR. Strict isolation with contact precautions, screening of patient contacts, co-horting of colonised and/or infected patients and dedicated nursing are advised. Treatment with two active drugs is recommended. Please contact your local microbiologist for further advice if needed.



## Pathology Report

Drs Du Buisson, Kramer, Swart, Bower Inc.  
Registration number: 2007/018337/21  
PR0520005200431

24hr Contact No. 011 709 1000

Doctor:

Dr T Law

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Batch: Email 436692 I-STD

3. Correlate with CXR

#### >AEROBIC CULTURE

#### BETA-LACTAMS

Ampi/Amox	R
Amox+Clav	R
Pip+Taz	R
Cefuroxime	R
Ceftriax/Cefota	R
Ceftazidime	R
Cef+Avi	S
Cefepime	R
Cef+Taz	R
Imipenem	R
Imipenem MIC 16	R
Meropenem	R
Meropenem MIC 32	R
Ertapenem	R
Doripenem	R
Doripenem MIC 32	R

#### AMINOGLYCOSIDES

Amikacin	S
Gentamicin	R

#### QUINOLONES

Ciprofloxacin	R
---------------	---

#### OTHER

Colist/Polymyx	S
Col/Poly MIC 1	S
Cotrimoxazole	R
Tigecycline	S
CARB	+

K. pneumo



New infiltrates on CXR  
Pathogen – needs treatment!






# 4. Molecular Testing of Respiratory Pathogens

	Core respiratory virus multiplex PCR	Comprehensive respiratory pathogens multiplex PCR	Triplex respiratory virus multiplex PCR
Components	<b>Viruses</b> SARS-CoV-2 Influenza A&B RSV A&B Adenovirus Metapneumovirus Parainfluenza 1-4 Rhinovirus	<b>Viruses</b> SARS-CoV-2 Influenza A&B RSV A&B Adenovirus Metapneumovirus Parainfluenza 1-4 Rhinovirus Enterovirus Coronavirus 229E Coronavirus NL63 Coronavirus OC43 Bocavirus <b>Bacteria</b> <i>B. pertussis</i> <i>B. parapertussis</i> <i>C. pneumoniae</i> <i>M. pneumoniae</i> <i>Legionella</i> spp.	<b>Viruses</b> SARS-CoV-2 Influenza A&B RSV A&B
Mnemonic	RVCORPCR	RPCOMPCR	COVTRIPCR


- Highly sensitive & specific viral PCRs have become the diagnostic “gold standard” in clinical virology
- Several of the newest assays also detect & identify the most common causes of “atypical” bacterial pneumonia
- NB: Not all positive results indicate current active infection**
- These results do not reflect on the microbiology cumulative reports as they are not from culture**




# Biofire® FilmArray® Pneumonia Panel plus

 Multiplexed PCR for simultaneous detection of multiple respiratory viral & bacterial pathogens in respiratory specimens (sputum, endotracheal aspirate & bronchoalveolar lavage)

- 18 bacteria
- 7 antibiotic resistance markers
- 9 viruses

 Run time: Approximately 1 hour from loading on the instrument

 **Utility of the panel is most relevant to ICU admissions**

 The 15 typical bacteria are semi-quantified to the nearest whole log as DNA copies/mL:  $10^4$ ,  $10^5$ ,  $10^6$ , and  $\geq 10^7$

- Can assist in distinguishing between an actual pathogen versus a coloniser

BIO  FIRE®  
BY BIOMÉRIEUX














 Few limitations:

- Nosocomial pathogens not covered: e.g. *S. maltophilia*, *Aspergillus* spp. & *Pneumocystis jirovecii*
- Detection of DNA doesn't imply the presence of a viable pathogen
- Susceptibilities of the organisms not known



# Biofire® FilmArray® Pneumonia Panel plus



Semi-Quantitative Bacteria	Atypical Bacteria	Antimicrobial Resistance Genes	
<p><b>Gram Negative Bacteria</b></p> <p> Enterobacteriales:</p> <ul style="list-style-type: none"> <li>▪ <i>Enterobacter cloacae</i> complex</li> <li>▪ <i>Escherichia coli</i></li> <li>▪ <i>Klebsiella pneumoniae</i> group</li> <li>▪ <i>Klebsiella oxytoca</i></li> <li>▪ <i>Klebsiella aerogenes</i></li> <li>▪ <i>Proteus</i> spp.</li> <li>▪ <i>Serratia marcescens</i></li> </ul> <p> Non-lactose fermenters:</p> <ul style="list-style-type: none"> <li>▪ <i>Acinetobacter calcoaceticus-baumannii</i> complex</li> <li>▪ <i>Pseudomonas aeruginosa</i></li> </ul> <p> <i>Haemophilus influenzae</i></p> <p> <i>Moraxella catarrhalis</i></p> <p><b>Gram Positive Bacteria</b></p> <ul style="list-style-type: none"> <li>▪ <i>Staphylococcus aureus</i></li> <li>▪ <i>Streptococcus agalactiae</i></li> <li>▪ <i>Streptococcus pneumoniae</i></li> <li>▪ <i>Streptococcus pyogenes</i></li> </ul>	<p><b>Atypical Bacteria</b></p> <ul style="list-style-type: none"> <li>▪ <i>Legionella pneumophila</i></li> <li>▪ <i>Mycoplasma pneumoniae</i></li> <li>▪ <i>Chlamydia pneumoniae</i></li> </ul> <p><b>Viruses</b></p> <ul style="list-style-type: none"> <li>▪ Influenza A</li> <li>▪ Influenza B</li> <li>▪ RSV</li> <li>▪ Human Rhinovirus/Enterovirus</li> <li>▪ Human Metapneumovirus</li> <li>▪ Parainfluenza virus</li> <li>▪ Adenovirus</li> <li>▪ Coronavirus (endemic)</li> <li>▪ MERS</li> </ul>	<p> Methicillin resistance:</p> <ul style="list-style-type: none"> <li>▪ <i>mecA/C &amp; MREJ</i>  <b>MRSA</b></li> </ul> <p> ESBL:</p> <ul style="list-style-type: none"> <li>▪ <i>CTX-M</i>  <b>ESBL</b></li> </ul> <p> Carbapenemases:</p> <ul style="list-style-type: none"> <li>▪ <i>OXA-48-like</i></li> <li>▪ <i>NDM</i></li> <li>▪ <i>VIM</i>  <b>CRE</b></li> <li>▪ <i>KPC</i></li> <li>▪ <i>IMP</i></li> </ul> <p style="text-align: center;"> BY BIOMÉRIEUX</p>	





# Molecular Testing of Other Respiratory Pathogens

■ Additional highly sensitive and specific PCRs are also available for testing of the following respiratory pathogens:

- *Pneumocystis jirovecii*
  - BD MAX: MSG multicopy gene
  - Sensitivity 98%, specificity 96%
- ***Mycobacterium tuberculosis* complex**
- ***Mycobacterium* spp. (NTM)**
- *Aspergillus* spp.
- CMV





## 5. Mycobacteria Diagnostic Tests

### Microscopy

- Conventional light microscopy (Ziehl Neelsen stain)
- LED fluorescent microscopy (Auramine O stain)

### Molecular testing

- Xpert MTB/RIF Ultra assay
- Direct TB PCR (pulmonary specimens): BD Max
- Direct TB PCR (non-pulmonary specimens): ELITE InGenius
- Hain Line Probe Assays (1<sup>st</sup>- and 2<sup>nd</sup>-line)
- Non-tuberculosis Mycobacteria PCR (Direct & culture)

### Culture

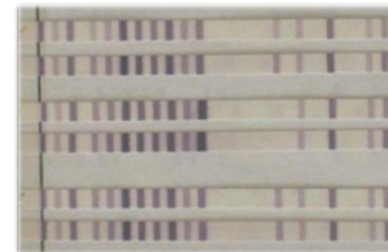
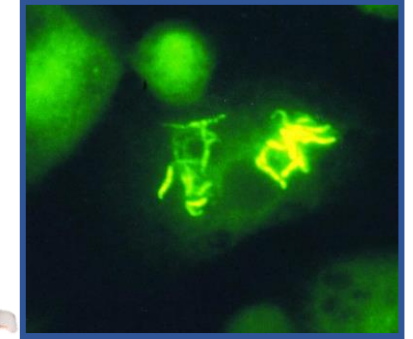
- Culture with specialised liquid culture systems  
(Mycobacteria Growth Indicator Tube (MGIT) & Myco/F Lytic)

### Drug susceptibility testing (DST)

- 1<sup>st</sup>-line agents & 2<sup>nd</sup>-line agents

### LF-LAM Urine test

### QuantiFERON-TB Gold – Latent TB











# MTB: Sensitivity & Limit of Detection

## WHY IS TB CULTURE STILL THE GOLD STANDARD?



	Microscopy	MTBDR <i>plus</i>	Xpert MTB/RIF	Xpert MTB/RIF Ultra	Culture
Limit-of-detection	5,000-10,000 bacilli/ml	≈160 bacilli/ml	≈114 bacilli/ml	16 bacilli/ml	10-100 bacilli/ml
			  		

Steingart KR, et al. Expert Rev Anti Infect Ther 2007;5:327-31.  
Parsons LM, et al. CMR 2011;24(2):314-50.



## 6. Blood Cultures

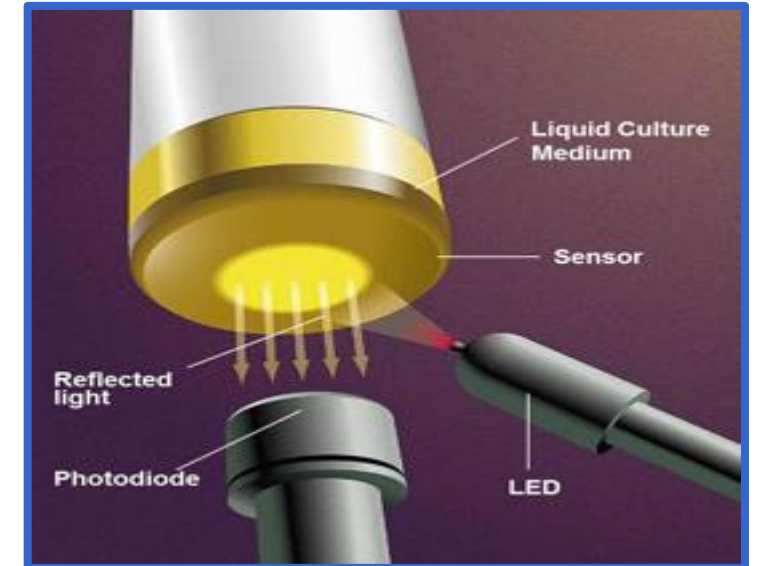
- **The major downside of doing blood cultures in patients with pneumonia, is their low diagnostic yield (positive cultures in 4.7-16%)**
- Ideally, send two sets of blood cultures for optimal yield (adults) in hospitalised patients only
- Blood cultures are incubated in automated, continuous-monitoring systems
- Once cultures become positive, an alarm & flashing light alert the lab staff
- A Gram stain is done immediately on a positive blood culture – this is useful in determining the possible aetiology of the bloodstream infection





# Blood Cultures

- CO<sub>2</sub> sensor bonded to bottom of blood culture bottle & separated from the broth by a semi-permeable membrane
- Sensor is impregnated with H<sub>2</sub>O vapour when bottles are autoclaved during manufacturing process
- Unidirectional membrane is permeable to CO<sub>2</sub> only
- Micro-organisms multiply in the media, generate CO<sub>2</sub> which diffuses across the membrane into the sensor & dissolves in the water, thereby generating H<sup>+</sup> ions
- Free H<sup>+</sup> ions interact with the sensor: **green (alkaline) → yellow (acidic)**
- As the sensor becomes lighter green & eventually yellow, it results in an increase of red light reflected by the sensor
- Once cultures become positive, the bottle is ejected from the Virtuo
- A Gram stain is done immediately – this is useful in determining the possible aetiology of the bloodstream infection





# Blood Culture Bottles

	Bottle Type	Purpose	Specimen Type
	BacT/ALERT SA	Standard Aerobic	Blood or SBF <i>SBF = Sterile body fluid</i>
	BacT/ALERT SN	Standard Anaerobic	Blood or SBF
	BacT/ALERT FA	FAN <sup>®</sup> Aerobic	Blood or SBF
	BacT/ALERT FN	FAN <sup>®</sup> Anaerobic	Blood or SBF
	BacT/ALERT PF	Paediatric FAN <sup>®</sup>	Blood only

Fill window and volume markers



For patient's on antibiotics

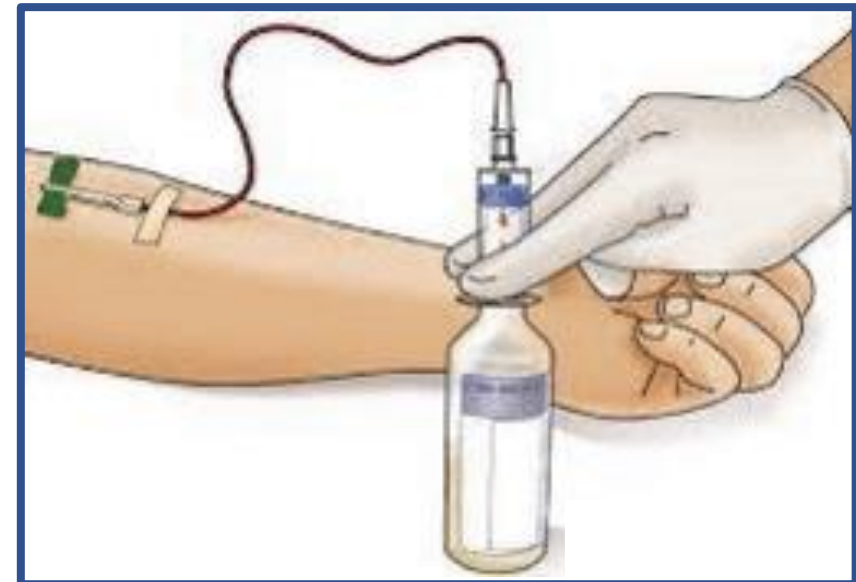
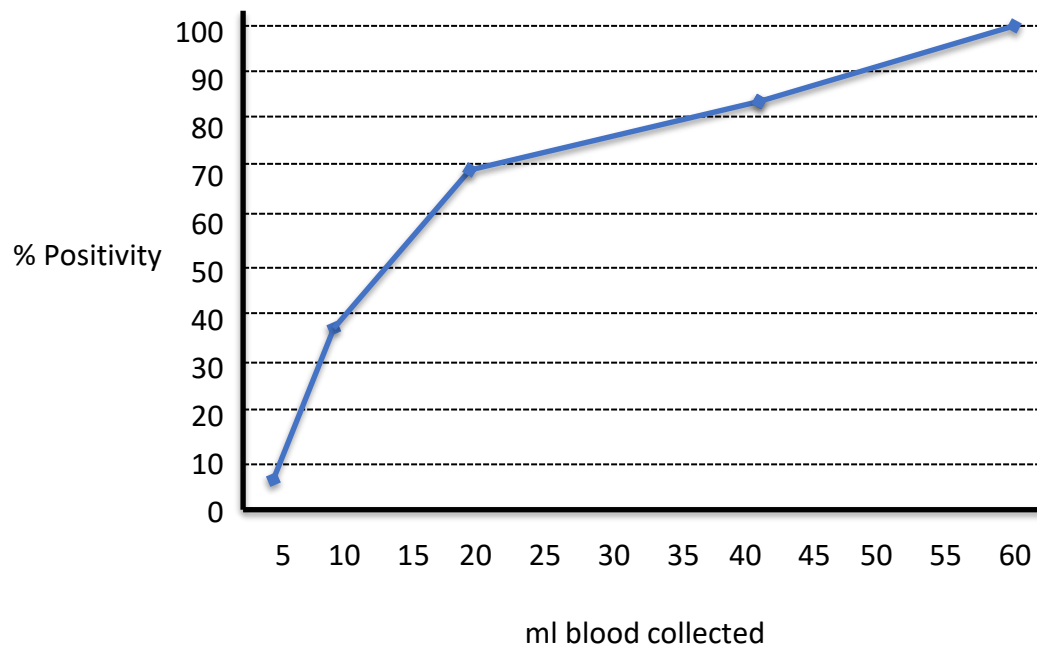
FAN<sup>®</sup> = Fastidious Antimicrobial Neutralization

- Adsorbent Polymeric Beads (APB) to neutralize antimicrobials



# Blood Cultures

- **The volume of blood obtained for each blood culture is the most important variable** in recovering pathogens from patients with bloodstream infections
- **Adults: 20 - 30 mL of blood per culture set is recommended - requires 2 - 3 bottles (i.e. 10mL per bottle)**
- *For adults it is always inappropriate to send a single yellow paediatric blood culture bottle*





# Blood Cultures

**Children:** Weight-appropriate volume of blood should be cultured

## Recommended Volumes of Blood for Culture in Paediatric Patients

Weight of patient	Culture set No. 1	Culture set No. 2	Total Volume for culture
≤ 1 kg	2 ml	-	2 ml
1.1 – 2 kg	2 ml	2 ml	4 ml
2.1 – 12.7 kg	4 ml	2 ml	6 ml
12.8 – 36.3 kg	10 ml	10 ml	20 ml
> 36.3 kg	20-30 ml	20-30 ml	40-60 ml

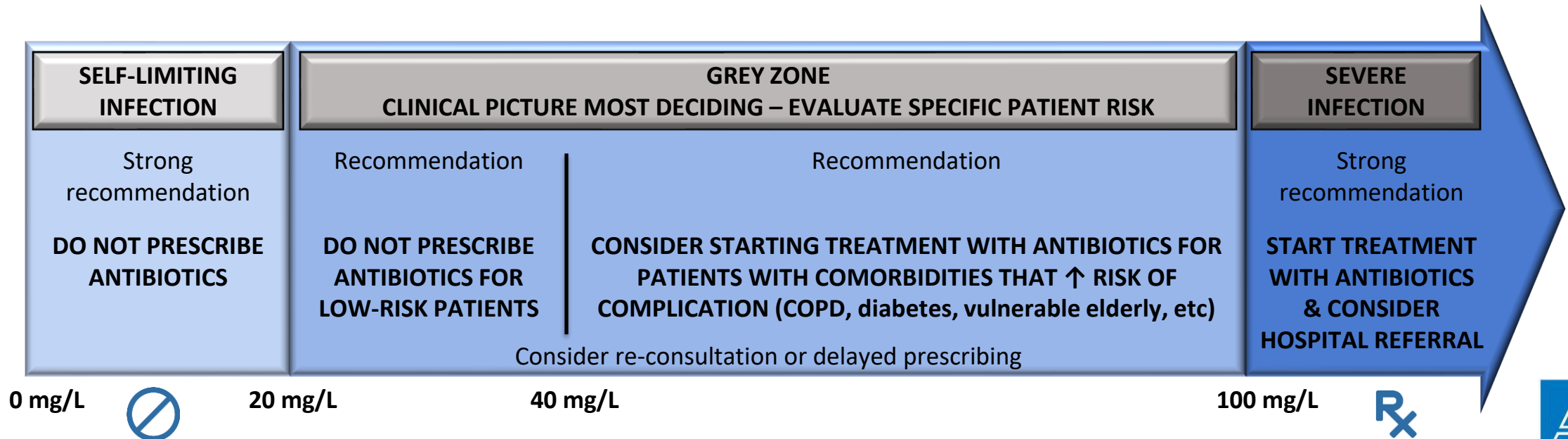
When ≤ 10 ml blood is collected, it should be inoculated into a single aerobic culture bottle





# 7. C-reactive protein (CRP)

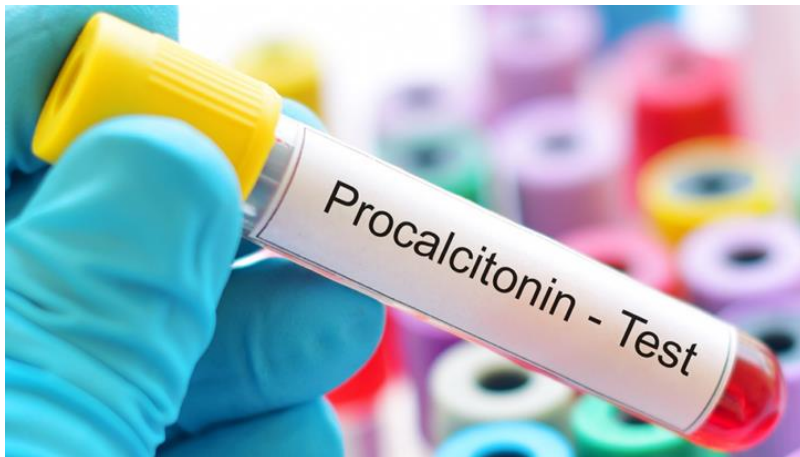
- A relatively **non-specific marker of inflammation** & therefore sepsis
- Levels are increased to some extent in most conditions associated with inflammation
- It is a cheap & widely available test & when levels are raised in a patient with signs suggestive of sepsis, it provides useful supporting evidence
- CRP increases slowly with a peak after 48-72 hours & a slow decrease thereafter





## 8. Procalcitonin (PCT)

- ▲ **PCT is a more reliable marker of sepsis than CRP**
- ▲ It is useful in distinguishing bacterial from other forms of infection
- ▲ PCT levels rise within 2-4 hours
- ▲ PCT kinetics then mirror the severity of infection
- ▲ **Evidence has shown that PCT is a useful method in guiding the initiation & duration of antibiotic treatment**
- ▲ PCT levels drop by  $\pm 50\%$  daily when infection is controlled & responds adequately to antibiotics



### PCT can be elevated by non-bacterial-infectious causes<sup>2</sup>

#### Physiologic stress

- Newborns (<48-72 hours; after 72 hours interpret levels as usual)
- Massive stress (severe trauma, surgery, cardiac shock, burns, pre-eclampsia)
- Prolonged, severe cardiogenic shock or organ perfusion abnormalities

#### Nonbacterial cytokine activation

- Some forms of vasculitis & acute graft vs. host disease
- Pancreatitis, chemical pneumonitis, mesenteric infarction, pulmonary aspiration
- Invasive fungal infections (e.g. candidiasis, aspergillosis) or acute *P. falciparum* malaria
- Chronic renal disease ( $\approx 2X$  increase in baseline levels), peritoneal dialysis or hemodialysis treatment
- Acute or chronic viral hepatitis, decompensated severe liver cirrhosis (Child-Pugh Class C) &/or acute liver failure
- Kawasaki, Still's Disease or Bell's Palsy

#### Dysregulated PCT production

- Treatment with agents that stimulate cytokines (OKT3, OK-432, TNF-alpha, antilymphocyte globulins, alemtuzumab, IL-2, granulocyte transfusion)
- Paraneoplastic syndromes due to medullary C-cell carcinoma, small cell lung carcinoma, or bronchial carcinoid

1. Noviello S, Huang DB. The basics and the advancements in diagnosis of bacterial lower respiratory tract infections. *Diagnostics* 2019;9:37.  
2. Samsudin I, et al. Clinical Utility and Measurement of Procalcitonin. *Clin Biochem Rev* 2017;38(2):59-68.



# How to Use PCT

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1 EVALUATE Determine if antibiotics are necessary	
< 0.1 ng/mL	Antibiotics are strongly discouraged
0.1-0.25 ng/mL	Antibiotics are discouraged
0.26-0.5 ng/mL	Antibiotics are encouraged
> 0.5 ng/ml	Antibiotics are strongly encouraged

2 MONITOR Assess therapy effectiveness over time
Test follow-up samples once every 1-2 days, to support decision to discontinue antibiotic therapy

3 DISCONTINUE Assess when to discontinue antibiotics	
≤ 0.25 ng/mL OR ΔPCT > 80%	Discontinuation of antibiotics is encouraged

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1 EVALUATE Assess sepsis risk & severity	
< 0.5 ng/mL	Low risk for progression to severe sepsis &/or septic shock
0.5-2.0 ng/mL	Sepsis should be considered
> 2.0ng/ml	High risk for progression to severe sepsis &/or septic shock

2 MONITOR Assess risk over time
Test follow-up samples once every 1-2 days, to support decision to discontinue antibiotic therapy

3 DISCONTINUE Assess when to discontinue antibiotics	
≤ 0.5 ng/mL OR ΔPCT > 80%	Discontinuation of antibiotics is encouraged



# In a nutshell



**Poor-quality specimens lead to misleading results, inappropriate antimicrobial therapy & delay in diagnosis**



**The quality of specimens is interpreted according to the microscopy & culture findings**



**Organisms from non-sterile sites may be colonisers & not necessarily the cause of infection**



**If the patient does not respond to antibiotic therapy chosen according to the microbiological susceptibility profile, consider:**

- inadequate therapy for the infection site
- inadequate dosage
- poor source control
- possible selection of resistant mutants
- different organism is causing the infection



# Communication



To determine significance of results requires knowledge of the patient's clinical status



The microbiology report should encourage communication between the clinician, pharmacist & the medical microbiologist



**Good communication makes a huge difference to patient management**



# Thank You

